

Nearly complete mitochondrial genome of *Trachelus stipa* Budak, Blank & Basibuyuk, 2017 (Hymenoptera: Cephidae)

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Abstract

The family Cephidae is a relatively modest phytophagous lineage within the suborder Symphyta of the order Hymenoptera. Members of this exclusively endophytophagous lineage feed on graminoids, berry canes, shrubs, and arboreal species. In this study, the nearly complete mitogenome of *Trachelus stipa* (Hymenoptera: Cephidae) was assembled and annotated as a third-party annotation and compared with the mitogenome of *T. iudaicus* and *T. tabidus*. The obtained length of the partial mitogenome of *T. stipa* was 16,085 bp with an A+T content of 79.02%. The mitogenome was containing 13 protein-coding, two rRNA, 22 tRNA genes and all of the protein coding genes (PCGs) are initiated with ATN as start codon, and all are ended with TAA or T– stop codon. All tRNA genes folded into the usual clover-leaf structure except for *tRNA-Ser1*. Compared with the inferred insect ancestral mitogenome, the gene orders are mostly conserved, except for the translocation of *tRNA-Met* and swapped positions of *tRNA-Ile* and *tRNA-Gln*. Phylogenetic analysis confirmed the position of *T. stipa* within genus *Trachelus* and recovered a relationship of *Stenocephus* + (*Janus* + ((*Phylloecus* + *Pachycephus*) + (*Syrista* + (*Calameuta* + (*Trachelus* + *Cephus*)))) in Cephidae.

Keywords: Cephidae phylogeny, mitogenome, *Trachelus*, Symphyta

1. Introduction

The family Cephidae constitutes a relatively modest phytophagous lineage within the Hymenoptera, comprising 165 described species across three subfamilies, of which Cephinae is the most diverse, encompassing 159 species distributed among 20 genera (Taeger et al., 2018). Members of this obligately endophytophagous lineage display varied host-plant associations and are conventionally referred as stem sawflies, reflecting the lifestyle of the larvae from a range of hosts, including graminoids, berry canes, shrubs, and arboreal species (Shanower & Hoelmer, 2004; Budak et al., 2011; Korkmaz et al., 2018). From an economic perspective, a number of species are known as important pests, such as larval stem borers inside grass stems or twigs of woody plants (Middlekauff, 1969; Shanower and Hoelmer, 2004).

The genus *Trachelus* Jurine, 1807, resides within the subfamily Cephinae and tribus Cephini, comprising nine described species primarily distributed across the Palearctic region, with one species (*T. tabidus*) introduced to North America since the 1880s (Taeger et al., 2018). The phylogenetic analyses confirm the monophyly of *Trachelus*, positioning it as closely allied to genera such as *Calameuta* and *Cephus*, within

a broader clade where traditional tribal divisions (Cephini, Hartigiini, Pachycephini) are not fully monophyletic (Budak et al., 2011; Korkmaz et al., 2018). The species of this genus exhibit specialized morphologies, including elongate antennae and genitalic structures adapted for oviposition in grass stems, with host associations predominantly involving Poaceae. Notably, *Trachelus stipa* Budak, Blank & Basibuyuk, 2017 is a recently described endemic species from Central Anatolia and displays morphological affinities to *T. flavicornis* and *T. troglodyta* in antennal segmentation and penile valve configuration, while specializing on *Stipa holosericea* (feather grass) as its larval host (Budak et al., 2017). Speciation dynamics in *Trachelus*, are most probably driven by host-plant transitions, as verified by phylogenetic reconstructions that highlight clade-specific adaptations to the host plants, with localized endemics such as *T. stipa* representing ecological speciation through niche adaptation within grassland biomes (Budak et al., 2011, 2017; Korkmaz et al., 2018).

Here, the nearly complete mitogenome of the stem sawfly *T. stipa* was assembled and annotated for the first time as a third-party annotation. So far, only two mitogenomes from *T. tabidus* and *T. iudaicus*, have been reported for this genus (Korkmaz et al., 2017). The mitogenome of *T. stipa* was compared with these previously reported mitogenomes of *Trachelus* to investigate the mitogenomic architecture and features within the genus.

2. Materials and Methods

Mitogenome Assembly and Annotation

The raw next generation sequencing (NGS) data of *T. stipa* was downloaded from the NCBI Sequence Read Archive (SRA) database with the accession number SRR27337724 using SRA Toolkit (<https://trace.ncbi.nlm.nih.gov/Traces/sra/sra.cgi?view=software>). The quality of the raw reads was evaluated using FastQC v0.12.1 (Andrews, 2010). Adapter sequences, low quality reads and possible reads including too much “N”s were eliminated from raw reads using fastp v0.20.0 (Chen et al., 2018). Clean reads were then assembled into mitogenomic contigs using MitoZ v3.6 (Meng et al., 2019) by using MEGAHIT (Li et al., 2015) as an assembler.

The annotation of the obtained mitogenome was performed using MitoZ v3.6 and MITOS2 (Al Arab et al., 2017; Donath et al., 2019) tool on Galaxy web server (Abueg et al., 2024). The decision on the locations and boundaries of the rRNA genes and PCGs was made manually by comparing with the *T. iudaicus* (NC032071) and *T. tabidus* (NC032072) mitochondrial genomes. The overlapping and intergenic regions between all genes were predicted manually. Finally, the nearly complete mitogenome of *T. stipa* was deposited in GenBank. The mitogenome is visualized using OrganellarGenomeDRAW (OGDRAW) (Greiner et al., 2019).

The nucleotide compositions were computed using MEGA v12.1 (Stecher et al., 2025). The strand asymmetries were calculated using the formulas: $AT\text{-skew} = [A - T]/[A + T]$ and $GC\text{-skew} = [G - C]/[G + C]$ (Perna and Kocher, 1995). To estimate the nucleotide and amino acid divergences between *Trachelus* species, the nucleotide and amino acid identities were calculated MEGA v12.1.

Phylogenetic analyses

Phylogenetic analyses were conducted on the mitogenome dataset of cephid species, including one newly annotated mitogenome of *T. stipa* (Table 1) to verify the phylogenetic position of *T. stipa*. *Allantus luctifer* (Tenthredinidae) was included as an outgroup (Table 1). The mitogenome sequences of the species that used in the phylogenetic analyses were retrieved from GenBank. Mitochondrial PCGs were aligned individually using “translation align” option implemented in Geneious R9 (Kearse et al., 2012) with MAFFT (Katoh and Standley, 2013) algorithm. Each rRNA gene was aligned individually using MAFFT implementation in Geneious R9. The individual alignments of each gene were then concatenated using PhyloSuite (Zhang et al., 2020). The optimal partition strategy and nucleotide substitution model of each partition were selected using ModelFinder (Kalyaanamoorthy et al., 2017) in PhyloSuite under the Bayesian Information Criterion (BIC). Best partitioning scheme and evolutionary model were used in subsequent phylogenetic analyses. The phylogenetic relationships were inferred with Maximum Likelihood (ML) approach. The ML analysis was carried out in IQ-TREE 2 (Minh et al., 2020) with 1000 bootstrap replicates using the proposed evolutionary model by PartitionFinder2. The produced phylogenetic trees were visualised by FigTree (Rambaut, 2014).

3. Results and Discussion

Mitochondrial genome architecture and nucleotide composition

The nearly complete mitogenome of *T. stipa* was assembled, annotated and compared with the mitogenome sequences of *T. iudaicus* and *T. tabidus* (Korkmaz et al., 2017). Sequence information for the part comprising the AT-rich region could not be obtained (Figure 1, Table 2). The length of the obtained mitogenome was 16,085 bp and including the 13 PCGs, 22 tRNAs and two rRNA genes (Figure 1, Table 2). The mitogenome organisation of the *T. stipa* was exactly the same with those of other reported *Trachelus* mitogenomes (Korkmaz et al., 2017). The mitogenome was mostly arranged in the same direction and order of the insect ancestral mitogenome, with only exception of the location of several tRNA genes, similar to previously reported cephid mitogenomes (Korkmaz et al., 2015; Korkmaz et al., 2016; Korkmaz et al., 2018). In total, twenty-three genes were encoded on the majority (J) strand, while the remaining fourteen genes were found on the minority (N) strand (Table 2).

The nucleotide composition of the mitogenome of *T. stipa* was remarkably biased towards A and T nucleotides, with a 79.02% A+T content, and similar to other reported *Trachelus* mitogenomes, varying between 79.03% A+T in *T. iudaicus* and 80.70% A+T in *T. tabidus* (Korkmaz et al., 2017) (Table 3). A similar bias towards A and T nucleotides was also detected in PCGs of the *T. stipa* mitogenome with 77.00% A+T content. The percentage of A+T of the 3rd codon position (86.18%) was substantially higher than those of the 1st (73.68%) and also the 2nd codon positions (71.14%) (Table 3). Similar to other reported *Trachelus* mitogenomes (Table 3), the AT- and GC- skews were found slightly positive (0.0750) and moderately negative (-0.2786) in the whole mitogenome of *T. stipa*, respectively.

Protein coding genes

The total length of the 13 PCGs was 11,240 bp which accounts for about 69.88% of the mitogenome (Table 2). Among the PCGs the *ND5* was the longest (1686 bp) and the *ATP8* was the shortest (177 bp). The lengths of PCGs were mostly similar to those of other *Trachelus* mitogenomes, except for *ND2*, *ATP8*, *COX3*, *ND4* and *ND6* (Table 2). The gene exhibiting the most length variation was the *COX3* with 9 codon differences between *T. stipa* (789 bp) and *T. iudaicus* (816 bp). As widely reported for the metazoan mitogenomes (Crozier and Crozier 1993), the PCGs was using the ATN-Ile/Met initiation codon. As frequently seen in invertebrate mitogenomes, TAA canonical termination codon was found in eleven PCGs (Table 2), while *ND2* and *COX1* have truncated termination codon (T–). The products of these two genes were possibly completed with the posttranscriptional polyadenylation process (Ojala et al. 1980, 1981). Among *Trachelus* species, based on both amino acid and nucleotide identities the least conserved PCG was *ATP8* (53.85% amino acid identity, 58.46% nucleotide identity), whereas the most conserved was *COX1* based on amino acid identity (94.52%), and was *ND5* based on nucleotide identity (82.29%) (Table 4).

tRNA and rRNA genes

All of the tRNA genes folded into usual clover-leaf secondary structure, except for *tRNA-Ser1*. Their locations and orientations are identical between *Trachelus* species, and the lengths were ranging from 65 bp (*tRNA-Arg* and *tRNA-Asn*) to 75 bp (*tRNA-His*). The nucleotide composition of the tRNA genes was biased towards A + T similar to mitogenome, with an average 84.14% A+ T.

The locations of rRNA genes were identical with those of other reported cephid mitogenomes (Figure 1) (Korkmaz et al., 2015; Korkmaz et al., 2016; Korkmaz et al., 2018; Liu et al., 2022). The *16S rRNA* was located between *tRNA-Leu1* and *tRNA-Val* and was 1396 bp in length (Table 2). The *12S rRNA* was found between *tRNA-Val* and *tRNA-Met* and its length was 1007 bp. The lengths of the rRNA genes were comparable with the reported *Trachelus* mitogenomes (*16S rRNA*: ranging between 1370 bp and 1406 bp, *12S rRNA*: ranging between 1021 bp and 1036 bp) (Table 2). The A+T nucleotide content of the rRNA genes was 83.98% on average in *T. stipa* mitogenome (Table 3).

Overlapping and noncoding regions

The total length of the overlapping regions between genes in the *T. stipa* mitogenome was 27 bp in six locations ranging between 1 and 8 bp (Table 2). The common reported “ATNNTAA/termination codon-N-initiation codon” overlap motif in insect mitogenomes was found in *ATP8-ATP6* and *ND4-ND4L* overlaps. The total length of intergenic regions was 261 bp and distributed in 18 different locations with a size ranging from 1 to 58 bp (Table 2). The longest intergenic region was located between *tRNA-Ile* and *ND2* genes (Table 2). There was also a non-coding region with a 585 bp length downstream of *tRNA-Gln*, which is possibly a part of AT-rich region, however the entire sequence of AT-rich region could not be obtained.

Phylogeny of Cephidae

The phylogenetic analysis has recovered the tree topologies with high bootstrap support values (Figure 2). The recovered tree confirmed the taxonomic position of *T. stipa* within genus *Trachelus* with high support (BS: 100%). The tree supported the monophyly of all included genera (Figure 2), and the topology was mostly congruent with the previously reported Cephidae and Hymenoptera phylogenies (Korkmaz et al., 2017; Korkmaz et al., 2018; Liston and Prous, 2021; Liu et al., 2022). The tree revealed sister group relationship of the genus *Cephus* and *Trachelus* (Figure 2). The genus *Calameuta* was found as sister to *Cephus* + *Trachelus* clade. In the Cephidae, a relationship of *Stenocephus* + (*Janus* + ((*Phylloecus* + *Pachycephus*) + (*Syrista* + (*Calameuta* + (*Trachelus* + *Cephus*)))) was recovered with high support values (Figure 2).

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Table 1. List of mitogenomes used in phylogenetic analysis

	Species	Family	Accession Number	References
	<i>Trachelus stipa</i>	Cephidae	Pending	This study
	<i>Trachelus iudaicus</i>	Cephidae	NC032071	Korkmaz et al., 2017
	<i>Trachelus tabidus</i>	Cephidae	NC032072	Korkmaz et al., 2017
	<i>Calameuta filiformis</i>	Cephidae	NC028445	Korkmaz et al., 2016
	<i>Calameuta idolon</i>	Cephidae	NC028446	Korkmaz et al., 2016
	<i>Cephus cinctus</i>	Cephidae	NC012688	Dowton et al., 2009
	<i>Cephus fumipennis</i>	Cephidae	PX255344	An, 2025
	<i>Cephus nigrinus</i>	Cephidae	BK071385	Hastaoglu Orgen, 2025
	<i>Cephus pygmeus</i>	Cephidae	KM377623	Korkmaz et al., 2015
Ingroup	<i>Cephus sareptanus</i>	Cephidae	KM377624	Korkmaz et al., 2015
	<i>Janus compressus</i>	Cephidae	KX907844	Korkmaz et al., 2018
	<i>Janus</i> sp.	Cephidae	OL757402	Niu and Wei, 2025
	<i>Janus</i> sp. 1 GYN-2022e	Cephidae	OL757400	Niu et al., 2025
	<i>Pachycephus cruentatus</i>	Cephidae	KX907845	Korkmaz et al., 2018
	<i>Pachycephus smyrnensis</i>	Cephidae	KX907846	Korkmaz et al., 2018
	<i>Phylloecus linearis</i>	Cephidae	KX907843	Korkmaz et al., 2018
	<i>Stenocephus fraxini</i>	Cephidae	NC069641	Niu and Wei, 2025
	<i>Syrista parreyssii</i>	Cephidae	OK104785	Liu et al., 2022
	<i>Syrista</i> sp. 1 GYN-2021c	Cephidae	OL889929	Niu, 2025
Outgroup	<i>Allantus luctifer</i>	Outgroup	NC024664	Wei et al., 2014

Table 2. Mitogenome summary of *Trachelus* species

Gene	Str and	<i>Trachelus stipa</i>						<i>Trachelus iudaicus</i>						<i>Trachelus tabidus</i>					
		Sta	En	Si	I	Start	Stop	Sta	En	Si	I	rt	p	Sta	En	Si	I	rt	p
		rt	d	ze	G	codon	codon	rt	d	ze	G	co	co	rt	d	ze	G	co	co
				(b	N							n	n					n	n
				p)															
<i>tRNA-Gln</i>	N	58	65					1	69	69	59			1	69	69	33		
<i>tRNA-Ile</i>	J	68	75					12	19	68	55			10	17	68	87		
		7	4	68	58			9	6					3	0				
<i>ND2</i>	J	81	18	10		ATT	T--	25	12	10	0	A	T--	25	12	10	0	AT	T--
		3	36	24	0			2	90	39		A		8	99	42		T	
<i>tRNA-Trp</i>	J	18	19					12	13	73	-8			13	13	69	-8		
		37	07	71	-8			91	63					00	68				
<i>tRNA-Cys</i>	N	19	19					13	14	69	70			13	14	71	1		
		00	73	74	16			56	24					61	31				
<i>tRNA-Tyr</i>	N	19	20					14	15	69	70			14	14	67	15		
		90	57	68	28			95	63					33	99				
<i>COX1</i>	J	20	36	15		ATG	T--	16	31	15	0	AT	T--	15	30	15	0	AT	T--
		86	19	34	0			34	67	34		G		15	48	34		G	
<i>tRNA-Leu2</i>	J	36	36					31	32	69	0			30	31	69	0		
		20	88	69	0			68	36					49	17				
<i>COX2</i>	J	36	43	68		ATG	TAA	32	39	68	11	AT	TA	31	38	68	15	AT	TA
		89	72	4	6			37	20	4		G	A	18	01	4		G	A
<i>tRNA-Lys</i>	J	43	44					39	40	72	5			38	38	72	10		
		79	50	72	6			32	03					17	88				
<i>tRNA-Asp</i>	J	44	45					40	40	74	0			38	39	70	0		
		57	26	70	0			09	82					99	68				
<i>ATP8</i>	J	45	47	17		ATT	TAA	40	42	18	-7	AT	TA	39	41	16	-7	AT	TA
		27	03	7	-7			83	65	3		T	A	69	36	8		T	A
<i>ATP6</i>	J	46	53	67		ATG	TAA	42	49	67	3	AT	TA	41	48	67	6	AT	TA
		97	71	5	13			59	33	5		G	A	30	04	5		G	A
<i>COX3</i>	J	53	61	78		ATG	TAA	49	57	81	75	AT	TA	48	56	79	8	AT	TA
		85	73	9	21			37	52	6		A	A	11	08	8		A	A
<i>tRNA-Gly</i>	J	61	62					58	58	70	0			56	56	69	0		
		95	63	69	0			28	97					17	85				
<i>ND3</i>	J	62	66	35		ATT	TAA	58	62	35	0	AT	T--	56	60	35	0	AT	TA
		64	17	4	2			98	49	2		C		86	38	3		T	-
<i>tRNA-Ala</i>	J	66	66					62	63	67	1			60	61	66	15		
		20	85	66	14			50	16					39	04				

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Table 3. Nucleotide composition of the mitogenomes of *Trachelus* species

	T	C	A	G	AT%	GC%	AT-skew	GC-skew
Whole genome								
<i>T. stipa</i>	36.55	13.41	42.47	7.57	79.02	20.98	0.0750	-0.2786
<i>T. tabidus</i>	38.21	12.18	42.50	7.12	80.70	19.30	0.0532	-0.2621
<i>T. iudaicus</i>	36.55	13.58	42.48	7.39	79.03	20.97	0.0750	-0.2952
Protein coding genes								
<i>T. stipa</i>	42.44	11.63	34.56	11.37	77.00	23.00	-0.1022	-0.0113
<i>T. tabidus</i>	42.73	11.47	34.71	11.09	77.44	22.56	-0.1036	-0.0166
<i>T. iudaicus</i>	41.43	12.74	34.00	11.84	75.42	24.58	-0.0985	-0.0369
Protein coding genes-1st								
<i>T. stipa</i>	35.48	10.60	38.21	15.72	73.68	26.32	0.0371	0.1943
<i>T. tabidus</i>	35.81	10.40	38.08	15.71	73.89	26.11	0.0307	0.2037
<i>T. iudaicus</i>	34.99	10.95	38.86	15.19	73.85	26.15	0.0523	0.1621
Protein coding genes-2nd								
<i>T. stipa</i>	49.32	16.63	21.82	12.24	71.14	28.86	-0.3865	-0.1521
<i>T. tabidus</i>	49.52	16.44	21.62	12.43	71.14	28.86	-0.3922	-0.1389
<i>T. iudaicus</i>	49.16	16.72	21.43	12.69	70.59	29.41	-0.3927	-0.1369
Protein coding genes-3rd								
<i>T. stipa</i>	42.52	7.66	43.67	6.16	86.18	13.82	0.0134	-0.1085
<i>T. tabidus</i>	42.88	7.56	44.43	5.13	87.30	12.70	0.0178	-0.1916
<i>T. iudaicus</i>	40.12	10.56	41.70	7.62	81.82	18.18	0.0192	-0.1613
rRNAs								
<i>T. stipa</i>	46.61	5.04	37.37	10.99	83.98	16.02	-0.1100	0.3714
<i>T. tabidus</i>	45.41	5.28	38.33	10.97	83.74	16.26	-0.0846	0.3501
<i>T. iudaicus</i>	46.08	5.62	36.01	12.30	82.08	17.92	-0.1227	0.3724
tRNAs								
<i>T. stipa</i>	41.91	6.64	42.24	9.21	84.14	15.86	0.0039	0.1618
<i>T. tabidus</i>	41.83	7.03	41.56	9.59	83.39	16.61	-0.0031	0.1542
<i>T. iudaicus</i>	40.28	7.38	42.52	9.82	82.79	17.21	0.0271	0.1418

Table 4. Nucleotide and amino acid identities of mitochondrial protein coding genes among *Trachelus* species

	Nueclotide identity (%)	Amino acid identity (%)
<i>ATP6</i>	79,91	88,39
<i>ATP8</i>	58,46	53,85
<i>COX1</i>	81,41	94,52
<i>COX2</i>	77,68	85,02
<i>COX3</i>	78,35	87,82
<i>CYTB</i>	78,1	89,45
<i>ND1</i>	78,77	84,08
<i>ND2</i>	72,57	67,14
<i>ND3</i>	75,21	80,34
<i>ND4</i>	74,87	80,04
<i>ND4L</i>	74,57	69,07
<i>ND5</i>	82,29	83,96
<i>ND6</i>	66,16	61,73

